

**Synthesis of 4-¹³C-4-Hydroperoxycyclophosphamide and
Alpha-¹³C-4-Hydroperoxycyclophosphamide**

Chorng-Kei Ho⁺ and Susan M. Ludeman^{+##}

⁺Department of Chemistry, The Catholic University of America, Washington, D.C.

20064, [#]Department of Chemistry, Trinity College, Washington, D.C. 20017,

^{*}To whom requests for reprints should be addressed at Catholic University.

SUMMARY

New synthetic routes were developed for incorporating ¹³C into the oxazaphosphorine ring and nitrogen mustard functionality of cyclophosphamide metabolites. 1,2-¹³C₂-Vinylbromide and ethylene oxide were used to synthesize 3,4-¹³C₂-3-butenyl N,N-bis(2-chloroethyl)phosphorodiamidate via the intermediacy of 3,4-¹³C₂-3-buten-1-ol. Ozonolysis of the phosphorodiamidate gave 4-¹³C-4-hydroperoxycyclophosphamide with an overall yield of 16% which was 27-times higher than a previously reported synthesis. As an extension of this synthetic pathway, 2-¹³C-ethyl bromoacetate was used to incorporate ¹³C into the C₁ position of bis(2-chloroethyl)amine hydrochloride which was used as a precursor to alpha-¹³C-4-hydroperoxycyclophosphamide (obtained in 16% overall yield). Also discussed are applications of these pathways to the synthesis of diversely labelled (¹⁴C, ¹³C, 2H) 4-hydroperoxycyclophosphamides, chirally-labelled oxazaphosphorines, phosphoramidate mustards, and related alkylating agents.

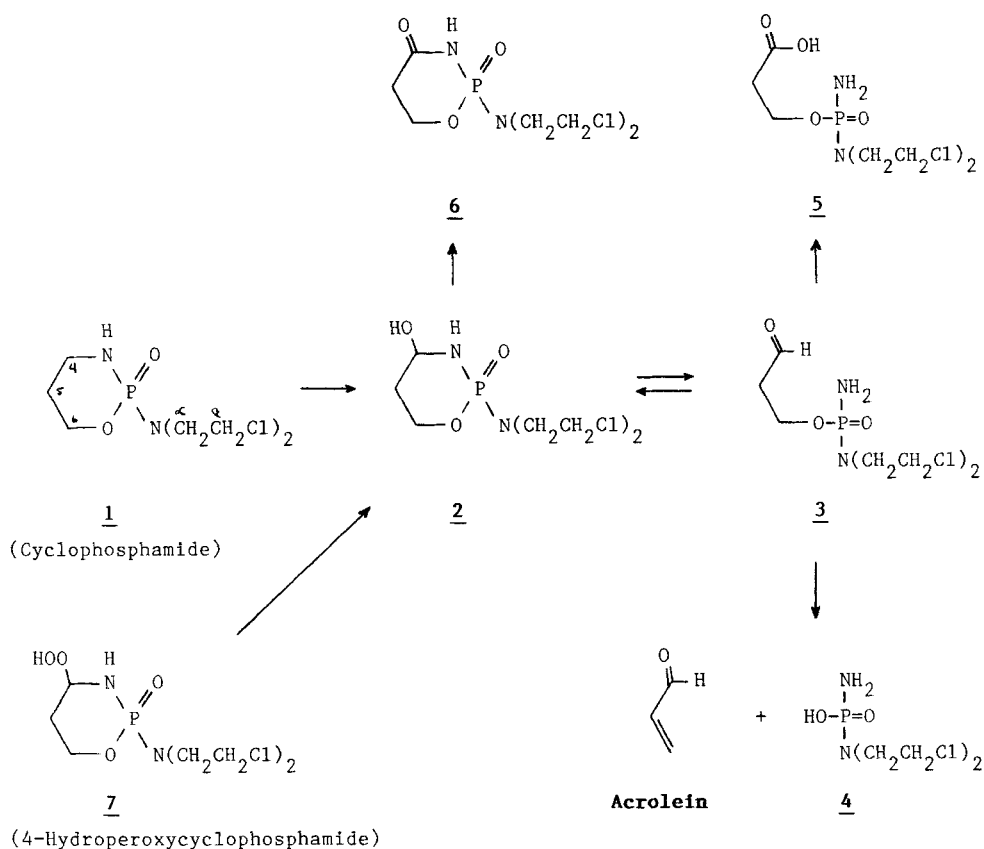
KEY WORDS: 4-¹³C-4-Hydroperoxycyclophosphamide; alpha-¹³C-4-hydroperoxycyclophosphamide; 3,4-¹³C₂-3-buten-1-ol; 1-¹³C-bis(2-chloroethyl)amine hydrochloride; labelled oxazaphosphorines.

INTRODUCTION

The therapeutic efficacy of the prodrug cyclophosphamide (**1**) in the treatment of many cancers is well established (1). As shown in Scheme I, the pertinent aspects of cyclophosphamide metabolism include an initial enzyme-mediated oxidation to give 4-hydroxycyclophosphamide (**2**) followed by a spontaneous ring-opening reaction to provide aldophosphamide (**3**). Metabolite **3** may undergo fragmentation to form acrolein and the ultimate alkylating agent phosphoramidate mustard (**4**) and/or **3** may be enzymatically oxidized to the non-toxic carboxyphosphamide (**5**). A second detoxification pathway is the enzymatic oxidation of **2** to ketocyclophosphamide (**6**) (2).

While the general features of the metabolism of 1 have been detailed, specific causal relationships between this chemistry and the oncostatic selectivity of 1 have not been established (2,3). NMR spectroscopy has been increasingly utilized as an analytical tool for the study of selectivity mechanisms as well as for related investigations of metabolite reactivity (5-10). ^{31}P NMR has proven extremely useful in this regard, but there are limitations (e.g. isochronous metabolite signals and the obvious inability to monitor metabolites which do not contain phosphorus) which can be overcome through the use of ^{13}C NMR and appropriately labelled metabolites.

Scheme I

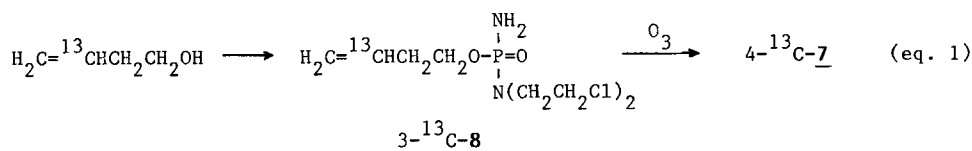


Access to the metabolites of 1 is conveniently provided by the reduction of 4-hydroperoxycyclophosphamide (7, Scheme I) (4,6); thus, incorporation of ^{13}C into 7 was the target of the work described herein. Among the various metabolites, the carbon atom which originates from the C_4 position in 7 is the most

distinctive; the ^{13}C NMR chemical shifts of this carbon in each of the metabolites (including various others not shown in Scheme I) have been reported (4,5). The unique spectroscopic character of this carbon provided the basis for our synthesis of 4- ^{13}C -4-hydroperoxycyclophosphamide (4- ^{13}C -7). For the purpose of monitoring the reactions of 4, alpha- ^{13}C -4-hydroperoxycyclophosphamide (alpha- ^{13}C -7) was also synthesized.

SYNTHESIS OF 4- ^{13}C -4-HYDROPEROXYCYCLOPHOSPHAMIDE

A synthesis of 4- ^{13}C -7 was previously reported (4); however, the overall yield of the multi-step route was only 0.6%. In our hands, the most effective synthesis of 7 involves the ozonolysis of 3-butenyl *N,N*-bis(2-chloroethyl)phosphorodiamidate (8), of which 3-buten-1-ol is a precursor (4). For the synthesis of 4- ^{13}C -7, 3- ^{13}C -3-buten-1-ol was thus a key intermediate to be prepared (eq. 1).

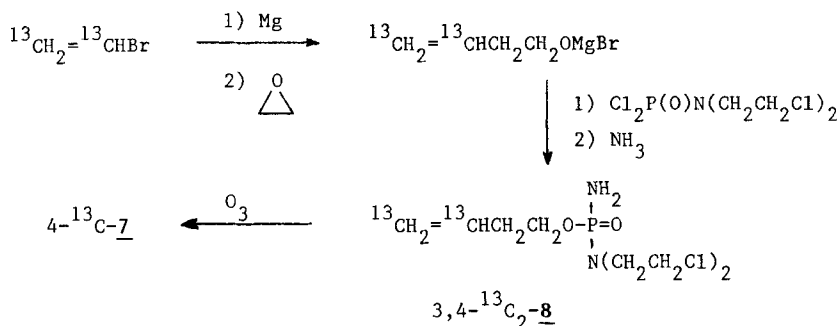


The initial synthetic scheme considered would have used ^{13}C -sodium cyanide as the source of the carbon label. In trial reactions, ethylene glycol was treated with sodium hydride followed by benzyl bromide to give 2-benzyloxyethanol (81%). Chlorination of this alcohol with thionyl chloride provided 2-benzyloxyethylchloride (92%) which was then converted to the iodide (78%) with sodium iodide. 2-Benzyloxyethyl iodide was reacted with sodium cyanide in DMSO to yield 3-benzyloxypropionitrile (91%). The nitrile was reduced with diisobutylaluminum hydride (DIBAL-H) and then hydrolysis with 10% H_2SO_4 (11) provided 3-benzyloxypropanal (53%). Multiple attempts to convert this aldehyde to 4-benzyloxy-1-butene (precursor to 3-buten-1-ol) via a Wittig reaction (12,13) were unsuccessful. This route was therefore abandoned in favor of a more direct synthesis of labelled 3-buten-1-ol, as outlined in Scheme II.

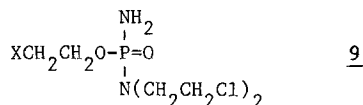
In initial trials using unlabelled materials, the intermediate 3-buten-1-ol (as generated by a Grignard reaction) was isolated by distillation (14,15). However, it was subsequently determined that 8 could be more efficiently synthesized through a 'one-pot' reaction which did not include the isolation of

the alcohol. On the other hand, the one-pot reaction led to a product mixture contaminated with a compound tentatively identified by ^1H and ^{13}C NMR as 9.

Scheme II



Formation of 9 could be accounted for by the reaction of ethylene oxide with halide(s) to give $\text{XCH}_2\text{CH}_2\text{O}^-$. In a variety of solvent systems, 8 and 9 had nearly identical R_f values which, therefore, necessitated multiple chromatographic separations in order to obtain pure 8 (attempts to ozonate crude mixtures of 8 and 9 led to poor recovery of hydroperoxide 7). The formation of 9 could be minimized by decreasing the amount of ethylene oxide and for our purposes it was determined that use of 0.75 equivalents of ethylene oxide (relative to starting material vinyl bromide) provided optimal conditions for generating 8 in a relatively efficient and cost-effective manner. By this method, a mixture of 8 and 9 was obtained in a ratio of 2:1, respectively. It can be noted that isolation of the intermediate 3-buten-1-ol obviated formation of 9 in subsequent reactions; however, the ultimate yield of 8 which was achieved was still lower than in the one-pot reaction. The yield of 8 also did not improve when vinyl lithium was prepared by substituting magnesium with lithium in Scheme II (16).

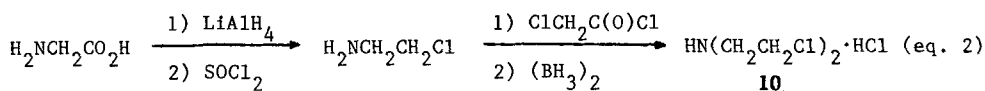


Incorporation of 1,2- $^{13}\text{C}_2$ -vinylbromide into Scheme II provided purified 3,4- $^{13}\text{C}_2$ -8 in 37% yield which was comparable to that achieved when unlabelled 8 was synthesized from commercially available 3-buten-1-ol (40%)(4). Ozonolysis of 3,4- $^{13}\text{C}_2$ -8 gave 4- ^{13}C -4-hydroperoxycyclophosphamide (4- ^{13}C -7) in 42% yield,

which was a marked improvement over yields reported for the synthesis of unlabelled 7 (27%)(4). The overall yield, then, of 4-¹³C-7 from 1,2-¹³C₂-vinylbromide was 16%, a 27-fold increase in efficiency over the previously reported synthesis of the same compound by an alternative pathway (4).

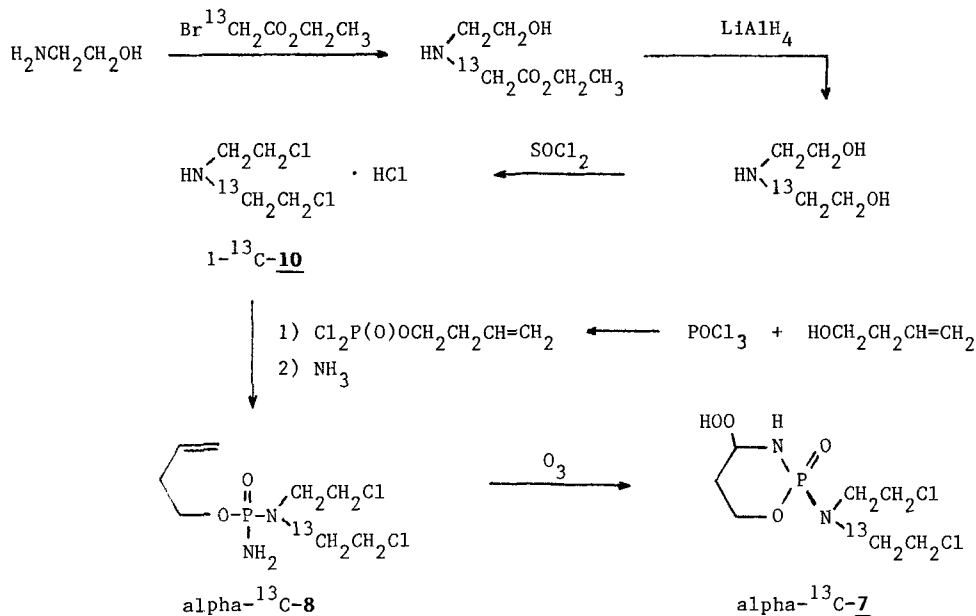
ALPHA-¹³C-4-HYDROPEROXYCYCLOPHOSPHAMIDE

For incorporation of ¹³C into the nitrogen mustard functionality of 7, the synthesis of 1-¹³C-bis(2-chloroethyl)amine hydrochloride (1-¹³C-10) was a key target. Since both glycine and chloroacetyl chloride labelled with ¹³C were commercially available, the synthesis of 10 was originally investigated as shown in eq. 2. However, this route gave various intermediates which were difficult to isolate and purify, and led to unacceptably low yields of 10.



Therefore, an alternative approach to the synthesis of 1-¹³C-10 and, ultimately, alpha-¹³C-7, was pursued as outlined in Scheme III.

SCHEME III



For the synthesis of butenyl phosphorodiamidate 8, typically 3-buten-1-ol is reacted with N,N-bis(2-chloroethyl)phosphorodiamidate dichloride (formed from

POCl_3 and 10), as shown in Scheme II. However, a more efficient use of labelled 10 was achieved by first synthesizing 3-butenyl dichlorophosphate and then reacting this intermediate with 10, followed by ammonia, to form 8. Thus, incorporation of 2- ^{13}C -ethyl bromoacetate into the synthesis shown in Scheme III provided α - ^{13}C -7 in an overall yield of 16%.

CONCLUSION

Schemes II and III outline specific syntheses of 4- ^{13}C -4-hydroperoxycyclophosphamide (4- ^{13}C -7) and α - ^{13}C -4-hydroperoxycyclophosphamide (α - ^{13}C -7), respectively. These synthetic pathways are very versatile, however, and could be applied to the synthesis of any number of variously labelled oxazaphosphorines and phosphoramidate mustards. For example, ethylene oxide is commercially available labelled with ^{14}C , ^{13}C , or ^2H and thus could be used to generate 7 with labels at the C_5 and C_6 positions. Incorporation of 2R,3R- or 2S,3S-2,3- $^2\text{H}_2$ -ethylene oxide (17,18) into Scheme II would yield 7 chirally-labelled at the C_5 and C_6 positions. Such chiral compounds would allow for stereochemical mechanistic studies of the fragmentation of 3 to acrolein and 4 (Scheme I).

Ethanolamine and ethyl bromoacetate (Scheme III) are commercially available or are readily synthesized from other compounds with ^{14}C , ^{13}C , or ^2H enrichment at various positions. Thus, ^{14}C and/or ^{13}C and/or ^2H could be incorporated into 10 at positions C_1 and/or C_2 and, furthermore, one or both of the chloroethyl groups could be labelled. These non-nitrogen mustards could then be used to synthesize diversely labelled phosphoramidate mustards, 4-hydroperoxycyclophosphamides, and related alkylating agents.

A particularly interesting extension of the syntheses described herein would be the synthesis of labelled 4-hydroperoxyifosfamide; ifosfamide is a clinically useful, structural isomer of cyclophosphamide which undergoes a similar metabolism. Substitution of $\text{ClP}(\text{O})(\text{HNCH}_2\text{CH}_2\text{Cl})_2$ for $\text{Cl}_2\text{P}(\text{O})\text{N}(\text{CH}_2\text{CH}_2\text{Cl})_2$ and ammonia in Scheme II would provide 4- ^{13}C -4-hydroperoxyifosfamide (19).

This work not only describes an improved synthesis of 4- ^{13}C -7 and a new synthesis of α - ^{13}C -7, but it also provides versatile synthetic pathways for making a wide variety of diversely labelled compounds related to the anticancer agent cyclophosphamide.

EXPERIMENTAL

Tetrahydrofuran (THF), Et₂O, benzene, P(O)Cl₃, and pyridine refer to freshly distilled solvents/reagents. Reaction mixtures that do not include water were carried out under nitrogen. Ozone (ca. 5–10 g/h) was produced by a Model 03V10-0 ozone generator (Ozone Research and Equipment Corp.). Analytical TLC employed 2.5 or 5 x 10 cm plates coated with a 250 micrometer layer of silica gel GF (Analtech). Flash chromatography employed 230–400 mesh silica gel (EM reagents). 89.55-MHz ¹H, 22.49-MHz ¹³C, and 36.23-MHz ³¹P NMR spectra were obtained with a JEOL FX-90Q broad-banded spectrometer. Unless specified otherwise, NMR samples were prepared using CDCl₃ and ¹H and ¹³C chemical shifts (ppm) refer to Me₄Si as an internal reference. ³¹P ppm values refer to external 25% H₃PO₄ in D₂O. In D₂O, ¹H chemical shifts refer to TSP as a reference.

3,4-¹³C₂-3-BUTENYL N,N-BIS(2-CHLOROETHYL)PHOSPHORODIAMIDATE (3,4-¹³C₂-8). THF (3.5 mL) and Mg (545 mg, 22.4 mmol) were added to a dry 50-mL 3-neck round-bottomed flask equipped with a liquid nitrogen condensor and septa. The mixture was cooled to -108 °C (THF/liquid N₂ bath) and then 1,2-¹³C₂-vinylbromide (2 g, 18.7 mmol, MSD Isotopes, 90 atom % ¹³C) was transferred as a gas to this flask from the manufacturer's glass vessel (which we equipped with a septum) via a double-ended needle using reduced pressure. The Grignard reaction was initiated by refluxing the reaction mixture (30 min); a spontaneous reflux was then allowed to continue for 10 min. An oil bath was then used to heat the reaction mixture at 85 °C for an additional 80 min. During the latter heating stage, THF was added at various stages (1 mL after 40 min and 5 mL after 50 min). After stirring at room temperature for 60 min, the reaction was cooled to -23 °C (CCl₄/dryice bath) and more THF (5 mL) was added followed by chilled ethylene oxide (576 mg, 13.1 mmol) in THF (5 mL). The mixture was stirred at -23 °C for 30 min, at 0 °C for 1.5 h, and at room temperature for 3 h. After cooling again to -23 °C, N,N-bis(2-chloroethyl)phosphoramidic dichloride (5.7 g, 22.4 mmol) in THF (18 mL) was added and the reaction stirred at low temperature for 2 h. After warming to 5 °C (ice bath), the reaction mixture was diluted with THF (10 mL) and NH₃ was then bubbled through the mixture for 15 min. The reaction flask was then stoppered and allowed to sit at room temperature overnight before filtration

and concentration at reduced pressure. The residual oil was flash chromatographed (3 cm x 50 cm column; CH₃OH - CH₂Cl₂, 1:50) giving 3.13 g of crude product (R_f 0.4 - 0.65, CH₃OH - CH₂Cl₂, 1:9). In ca. 1 g portions, this material was then repeatedly (2-3 times) chromatographed under pressure (20 psi) on silica gel (less than 230 mesh, EM Reagents, 3 cm x 50 cm column, CH₃OH - CH₂Cl₂, 1:50). Pure 3,4-¹³C₂-8 (R_f 0.5, CH₃OH - CH₂Cl₂, 1:9) was obtained as a colorless oil in 37% yield (1.93 g, 7 mmol) based on the amount of 1,2-¹³C₂-vinylbromide used in the initial reaction. ¹H NMR (ppm): 6.9 - 4.6 (doubled m, $^1J_{CH}$ = 151 Hz, 1H, ¹³CH=), 5.13 - 4.12 (doubled m, $^1J_{CH}$ = 154 Hz, 2H, ¹³CH₂=), 4.17 - 3.90 (m, 2H, CH₂O), 3.77 - 3.09 (m, 10 H, NH₂ and 2 NCH₂CH₂), and 2.4 (apparent q, J = 6.6 Hz, 2H, CH₂CH₂O). ¹³C NMR (ppm): 133.8 (d, $^1J_{CC}$ = 69.6 Hz, ¹³CH=) and 117.2 (d, $^1J_{CC}$ = 69.6 Hz, ¹³CH₂=). ³¹P NMR (ppm): 15.62.

4-¹³C-4-HYDROPEROXYCYCLOPHOSPHAMIDE (4-¹³C-7). Ozone was bubbled through a solution of 3,4-¹³C₂-8 (663 mg, 2.4 mmol) in acetone-water (2:1, 18 mL) at 5 °C (ice bath) for 15 min. The volume of the solution was adjusted to 18 mL with more acetone, aqueous H₂O₂ (0.90 mL of a 30% solution) was added, and the reaction flask was then stoppered and kept at room temperature overnight. The reaction mixture was concentrated on a rotary evaporator at ambient temperature and the residual aqueous solution was extracted with CH₂Cl₂ (6 x 25 mL). The combined extracts were dried (MgSO₄), concentrated at ambient temperature, and the residual material was then dissolved in minimal Et₂O and stored overnight at -20 °C. The crystalline product was separated and the mother liquor was purified by flash chromatography (2 cm x 15 cm column, CH₃OH - CH₂Cl₂, 4:96) and the fractions containing desired product (R_f 0.30) were concentrated; the residue was taken up in minimal Et₂O for crystallization at -20 °C. The combined yield of 4-¹³C-7 as a white microcrystalline solid was 42% (297 mg, 1 mmol). The ¹H NMR spectrum was consistent with cis stereochemistry (4, 20, 21). ¹H NMR (ppm): 5.15 (doubled, doubled q, $^1J_{CH}$ = 170 Hz, $^3J_{HH}$ = 2 Hz, $^3J_{HP}$ = 26 Hz, 1 H, ¹³C-H), 5.10 - 3.90 (m, 3H, OOH and CH₂O), 3.80 - 3.20 (m, 8H, 2 NCH₂CH₂Cl), and 2.20 - 1.91 (m, 3H, NH and CH₂CH₂O). ¹³C NMR (ppm): 86.45 (d, $^2J_{CP}$ = 3.7 Hz). ³¹P NMR (ppm): 9.91 (d, $^2J_{CP}$ = 3.7 Hz).

2-¹³C-ETHYL 2-(2-HYDROXYETHYLAMINO)ACETATE. 2-¹³C-Ethyl bromoacetate (2 g, 11.8 mmol, MSD Isotopes, 90 atom % ¹³C) was added dropwise to a solution of ethanolamine (2.18 g, 35.7 mmol) in CHCl₃ (12 mL). After stirring at room temperature for 20 min, the reaction mixture was concentrated and purified by flash chromatography (5 cm x 15 cm column, CH₃OH - CHCl₃, 8:92) giving the product as a colorless oil (R_f 0.35, 1.23 g, 8.3 mmol, 70% yield). ¹H NMR (ppm): 4.20 (q, J = 7.3 Hz, 2H, CO₂CH₂CH₃), 3.66 (t, J = 5.9 Hz, 2H, CH₂OH), 3.43 (d, $^1J_{CH}$ = 136.7 Hz, 2H, ¹³CH₂CO₂CH₂), 2.80 (t, J = 5.9 Hz, 2H, CH₂CH₂OH), 2.42 (broad s, 2H, NH and OH), and 1.29 (t, J = 7.3 Hz, 3H, CH₃). ¹³C NMR (ppm): 50.1.

1-¹³C-2,2'-IMINODIETHANOL. 2-¹³C-Ethyl 2-(2-hydroxyethylamino)acetate (1.20 g, 8.1 mmol) in THF (15 mL) was added dropwise to an ice-cooled suspension of LiAlH₄ (459 mg, 12.1 mmol) in THF (13 mL). Following complete addition, the reaction mixture was refluxed overnight and then cooled to room temperature. Following the sequential addition of H₂O (0.2 mL), 15% NaOH (0.2 mL), H₂O (0.8 mL), and THF (50 mL), the reaction mixture stirred overnight at room temperature. Filtration followed by concentration gave the product as a colorless oil which was used without further purification (737 mg, 6.9 mmol, 85%). ¹H NMR (ppm): 3.80 (s, 3H, NH and 2 OH), 3.68 (t, J = 5.9 Hz, 4H, two CH₂O), 2.73 (t, J = 5.9 Hz, 2H, NCH₂), and 2.73 (doubled t, $^1J_{CH}$ = 131.8 Hz, $^3J_{HH}$ = 5.9 Hz, 2H, N¹³CH₂). ¹³C NMR (ppm): 51.1.

1-¹³C-N,N-BIS(2-CHLOROETHYL)AMINE HYDROCHLORIDE (1-¹³C-10). A solution of 1-¹³C-2,2'-iminodiethanol (737 mg, 6.9 mmol) and thionyl chloride (4.14 g, 34.8 mmol) in CHCl₃ (8 mL) stirred at room temperature overnight and was then refluxed for 4 h. The reaction mixture was concentrated and the residual solid was purified by crystallization using Et₂O. Product 1-¹³C-10 was obtained in 81% yield (1.0 g). ¹H NMR (D₂O) (ppm): 3.82 (t, J = 5.9 Hz, 4H, 2 CH₂O), 3.43 (t, J = 5.9 Hz, 2H, NCH₂), and 3.43 (doubled t, $^1J_{CH}$ = 147 Hz, $^3J_{HH}$ = 5.9 Hz, 2H, N¹³CH₂).

3-BUTENYLPHOSPHORYL DICHLORIDE. POCl₃ (3.06 g, 20 mmol) was added dropwise to an ice-cooled solution of 3-buten-1-ol (1.73 mL, 20 mmol) in benzene (20 mL). This was followed by the dropwise addition of pyridine (1.58 g, 20 mmol). The reaction mixture stirred overnight at room temperature and was then filtered and

concentrated. The residue was purified by flash chromatography (4 cm x 15 cm column, CHCl_3) and the product was obtained as a colorless oil (R_f 0.70, 2.55 g, 13.5 mmol, 68%). ^1H NMR (ppm): 5.05 - 4.56 (m, 1 H, $\text{CH}_2=\text{CH}$), 4.23 - 4.04 (m, 2H, $\text{CH}_2=$), 3.52 - 3.21 (m, 2H, CH_2O), 3.20 - 2.40 (m, 2H, $\text{CH}_2\text{CH}_2\text{O}$). ^{31}P NMR (ppm): 6.87.

ALPHA- ^{13}C -3-BUTENYL *N,N*-BIS(2-CHLOROETHYL)PHOSPHORODIAMIDATE (ALPHA- ^{13}C -8).

Pyridine (569 mg, 5.6 mmol) was added dropwise to a suspension of 1- ^{13}C -10 (505 mg, 2.8 mmol) in CH_2Cl_2 (10 mL) at ice-bath temperature. This was followed (after ca. 5 min) by the dropwise addition of 3-butenylphosphoryl dichloride (1.57 g, 8.3 mmol) in CH_2Cl_2 (2 mL). After stirring at room temperature for 4 days, the reaction mixture was filtered, concentrated, and purified by flash chromatography (1.5 cm x 15 cm column, Et_2O). Fractions containing the desired intermediate (R_f 0.80) were concentrated at ambient temperature and then immediately diluted with THF (30 mL) and cooled to 5 $^\circ\text{C}$. NH_3 was bubbled through this cooled solution for 15 min and the reaction flask was then stoppered and allowed to stand overnight at room temperature. The reaction mixture was filtered, concentrated, and purified by flash chromatography (2 cm x 15 cm column, $\text{CH}_3\text{OH} - \text{CHCl}_3$, 4:96). Alpha- ^{13}C -8 was recovered as an oil (R_f 0.30, 602 mg, 2.2 mmol, 78%). ^1H NMR (ppm): 6.03 - 5.58 (m, 1 H, $\text{CH}_2=\text{CH}$), 5.28 - 5.00 (m, 2H, $\text{CH}_2=$), 4.04 (doubled, doubled triplet, $J_{\text{HH}} = 6.8$ Hz and 1.2 Hz, $^3J_{\text{HP}} = 6.8$ Hz, 2H, CH_2O), 3.77 - 3.30 (m, 6H, NCH_2 and 2 CH_2Cl), 3.51 (doubled, doubled triplet, $^1J_{\text{CH}} = 140.6$ Hz, $^3J_{\text{HP}} = 11.5$ Hz, $^3J_{\text{HH}} = 6.8$ Hz, 2H, N^{13}CH_2), 2.85 (broad s, 2H, NH_2), and 2.4 (apparent q, $J = 6.6$ Hz, 2H, $\text{CH}_2\text{CH}_2\text{O}$). ^{13}C NMR (ppm): 49.31 (d, $^2J_{\text{CP}} = 4.9$ Hz). ^{31}P NMR (ppm): 15.62 (d, $^2J_{\text{CP}} = 4.9$ Hz).

ALPHA- ^{13}C -4-HYDROPEROXYCYCLOPHOSPHAMIDE (ALPHA- ^{13}C -7). The title compound was prepared from alpha- ^{13}C -8 (594 mg, 2.2 mmol) using the same procedure described above for the synthesis of 4- ^{13}C -7. Alpha- ^{13}C -7 was obtained as a white micro-crystalline solid in 43% yield (274 mg, 0.93 mmol) with *cis* stereochemistry (4, 20, 21). ^1H NMR (ppm): 5.15 (doubled q, $J_{\text{HH}} = 2$ Hz, $^3J_{\text{HP}} = 26$ Hz, 1 H, H-COOH), 5.10 - 3.90 (m, 3H, OOH and CH_2O), 3.80 - 3.20 (m, 6H, NCH_2 and 2 CH_2Cl), 3.32 (doubled, doubled triplet, $^1J_{\text{CH}} = 135$ Hz, $^3J_{\text{HH}} = 6.8$ Hz, $^3J_{\text{HP}} = 12.2$ Hz, 2H,

N^{13}CH_2), and 2.20 - 1.91 (m, 3H, NH and $\text{CH}_2\text{CH}_2\text{O}$). ^{13}C NMR (ppm): 48.72 (d, $^2\text{J}_{\text{CP}} = 4.9$ Hz). ^{31}P NMR (ppm): 9.86 (d, $^2\text{J}_{\text{CP}} = 4.9$ Hz).

ACKNOWLEDGMENT

This investigation was supported by PHS Grant No. CA37323 (to S.M.L.) awarded by the National Cancer Institute, DHHS.

REFERENCES

1. Hill D.L. - A Review of Cyclophosphamide, Charles C. Thomas, Springfield, IL, 1975.
2. Friedman O.M., Myles A., and Colvin M. - Adv. Cancer Chemother. 1: 143 (1979).
3. Sladek N.E. - Metabolism and Action of Anticancer Drugs, Taylor and Francis, London, pp. 48-90, 1987.
4. Zon G., Ludeman S.M., Brandt J.A., Boyd V.L., Ozkan G., Egan W., and Shao K.-L. - J. Med. Chem. 27: 466 (1984).
5. Boyd V.L., Summers M.F., Ludeman S.M., Egan W., Zon G., and Regan J.B. - J. Med. Chem. 30: 366 (1987).
6. Boyd V.L., Robbins J.D., Egan W., and Ludeman S.M. - J. Med. Chem. 29: 1206 (1986).
7. Ludeman S.M., Egan W., Zon G., and Boal J.H. - NMR Spectroscopy in Drug Research, Munksgaard, Denmark, pp. 488-507, 1988.
8. Borch R.F., Hoyer T.R., and Swanson T.A. - J. Med. Chem. 27: 490 (1984).
9. Kwon C.H., Borch R.F., Engel J., and Niemeyer U. - J. Med. Chem. 30: 395 (1987).
10. Borch R.F. and Millard J.A. - J. Med. Chem. 30: 427 (1987).
11. Trofimenko S. - J. Org. Chem. 29: 3046 (1964).
12. Benson R.E. and McKusick B.C. - Organic Synthesis, Coll. Vol. 4, John Wiley and Sons, NY, pp. 746-752, 1963; Kogerman P.N. - J. Amer. Chem. Soc. 52: 5060 (1930).
13. Brown J.M., Golding B.T., and Stofko J.J. Jr. - J. Chem. Soc. Perkin II: 436 (1978).
14. Seyferth D. - Org. Syn., Coll. Vol., 4: 258 (1963).
15. Dreger E.E. - Org. Syn., Coll. Vol., 1 (second ed.): 306 (1946).
16. West R. and Glaze W.H. - J. Org. Chem. 26: 2096 (1961).
17. Schwab J.M. and Ho C.-K. - J. Chem. Soc., Chem. Commun.: 872 (1986).
18. Schwab J.M., Ray T., and Ho C.-K. - J. Amer. Chem. Soc. 111: 1057 (1989).
19. Boal J.H., Williamson M., Boyd V.L., Ludeman S.M., and Egan W. - J. Med. Chem. 32: 1768 (1989).

20. Takamizawa A., Matsumoto S., Iwata T., and Makino I. - *Heterocycles* 7: 1091 (1977).
21. Camerman A., Smith H.W., and Camerman N. - *Biochem. Biophys. Res. Commun.* 65: 828 (1975).